

Review Article

PARASITES AND CANCER: A REVIEW OF THE EMERGENCE OF PROTOZOAN CARCINOGENESIS AND NOVEL MOLECULAR INSIGHTS.

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ABSTRACT

The role of infectious agents in the formation of cancers has been long established. However the bulk of the emphasis has been on oncogenic DNA viruses and to a lesser extent, bacteria. However, amidst parasites, only a few metazoans have been linked to cancer, and with feeble molecular bases. This review explores the role of protozoa in cancer formation and highlights new insights into the process of oncogenesis by previously identified helminths with carcinogenic potential. It expounds on the impact of parasites on aspects of cell growth and function, particularly apoptosis, gene expression and cell proliferation.

KEYWORDS: Protozoa, Cancer, Oncogenesis, Helminths, Apoptosis.

NigerJMed2017: 82-88

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BACKGROUND

Cancer has assumed and maintained a state of public health importance. Unravelling the aetiopathogenesis of common cancers is key to effective control. Microbial infections have become increasingly important in the formation of cancers, and are of greater importance in tropical parts of the world. Most of the emphasis has been on viruses and bacteria, and although largely overlooked, protozoa are an increasingly important factor in microbial carcinogenesis.

The International Agency for Research on Cancer [IARC], in 1990, stated that *Schistosoma haematobium* infection was associated with a five-fold increase in the risk of squamous cell carcinoma of the bladder, while in 1994, direct links were documented between Liver Flukes [*Opisthorchis* sp and *Clonorchis* sp] and cholangiocarcinoma of both intra and extra hepatic bile ducts.^[1]

Notably *Plasmodium falciparum* infection has been labelled as a co-factor in the pathogenesis of endemic Burkitt lymphoma.^[2] Although several studies suggest an association between malaria and BL, there has never

been a conclusive population study in support of a direct role of malaria in causation of BL.^[2] Over the last few decades there has been an increased intensity in the search for scientifically viable links between parasites and cancers, especially in the background of “double burden of disease” in tropical regions as well as improved survival of immunosuppressed individuals. Cancers arise from abnormal unregulated growth of cells leading to abnormal form or function of body organs. It has attained a status of global public health importance, accounting for more deaths than HIV and Tuberculosis combined.^[3] Cancer has defied previously imposed labels of “disease of rich nations” accounting for an increasing number of deaths in Africa and other 3rd world regions. The cancer burden in developing countries is approaching pandemic proportions.^[3] More than half of the 12.4 million new cases of cancer in 2008, and two-thirds of the estimated 7.6 million deaths occur in low and middle income countries.^[3] Once shrouded in mystery, there has been an explosion of knowledge with regards the inception and progression of cancers. Consequently, the role of infection has been radically redefined in the last few years, attaining a state of increasing relevance. The American Cancer Society recently stated that infections are linked to 15-20% of cancers, while El-Gayar observed that cancer cases in developing countries can be lowered by more than 25%, if infections are properly treated.^[4,5] It is equally estimated that cancers attributable to infections

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accounted for 17.8% of global cancer burden, while there would be 7.7% less cancer cases in the developed world if associated infections were treated or prevented.^[6] Notably, it has been documented that cancers due to infections are progressively increasing.^[7]

Infectious agents have been known to incite a chronic inflammatory response with generation of genotoxic free radicals, an altered internal milieu of cytokines stimulating rapid cell turnover and metaplastic change.^[8] These create a biological setting predisposing to genetic damage and impaired DNA repair. However, microbial pathogens are now known to directly cause DNA mutations, cell cycle modulation, dysregulation of DNA and resistance to apoptosis.^[9] A larger part of the available literature on infection associated cancers has revolved around the role of viruses and bacteria. Insomuch that it has been once stated that “the classic theory of carcinogenesis revolves around radiation, chemicals and viral infection”.^[10] Helminths, on the other hand, are largely believed to largely cause self limiting infectious ailments, save in states of immunosuppression where the prognosis is worsened by an impaired immune response. Amidst parasites/helminths, only multicellular metazoans such as liver flukes and Schistosomes are presently recognized by the International Agency for Research in Cancer [IARC] to have established roles in carcinogenesis.^[1]

The first link between parasites and cancers was first highlighted by Fibiger in 1926.^[11] He observed that mice infected with *Spiroptera* later developed gastric cancer. This was later disproved by other researchers.^[11] However, this heightened the interest in the possible role of protozoa in cancers and has led to the unearthing of numerous new concepts about the impact of protozoa in the formation and progression of cancers. Recent research has led to an increase in the number of protozoan pathogens with suspected or confirmed roles as carcinogens. In the same vein, molecular changes that underlie the neoplastic changes seen in a background of known parasitic carcinogens have been unravelled a step further. This article will look at new molecular pathways implicated in parasite induced carcinogenesis, particularly with regards cell cycle regulation, apoptosis, and DNA repair, while exploring links between recently unassociated protozoans and cancers. It equally seeks to strengthen the case for increased surveillance, prophylaxis and treatment of parasitic infection as part of cancer control and prevention.

METHOD OF REVIEW

To review relevant literature on the role of parasites in the formation of cancers, emphasizing largely

overlooked protozoa and recently elucidated molecular pathways in both protozoan and metazoan carcinogenesis, we conducted a database search (on Pub Med and Google Scholar) using broad terms related to the intended review such as Parasites and Cancer, Carcinogenic Protozoa, Molecular basis of Parasitic Carcinogenesis. Etc. references in the identified articles were also screened. A total of 79 articles published in English were found, of which 11 were unobtainable. Articles found were screened for content and relevance. Of the 68 articles identified, 65 were found relevant and referenced.

PROTOZOA AND THEIR NOVEL PATHWAYS OF TUMORIGENESIS

Cryptosporidium parvum

This unicellular protozoan is a genus of Apicomplexa and class Sporozoa that causes infections of the gastrointestinal and respiratory tracts.^[12] It's a frequent cause of diarrhea in both immunocompetent and immune deficient individuals. Its presence however, carries a higher risk of colorectal malignancy in severe immunosuppression.^[13] A study in Poland showed that *Cryptosporidium parvum* was seen in 18% of cases of colorectal cancer, while a study by El-gayar observed that in mice having cryptosporidiosis, adenomas and invasive neoplasia were detected in the stomach duodenum and ileocecal region.^[5,14] A study in Egypt observed dysplastic changes in 80% of colonic biopsies of rats infected with *Cryptosporidium parvum*, while Certad documented an increase in mitosis, alongside low and high grade intra epithelial neoplasia in different areas of the digestive tract.^[14,15]

Increased mitosis was demonstrated on a molecular level in studies by Certad, which show enhanced staining with Ki-67, [a marker of mitosis] while an increased expression of the cell cycle protein, Cyclin D1 was observed in an animal study by Abdou et al.^[14,15] Several other studies have documented alterations in the expression of inhibitors of apoptosis such as Bcl2, as well as activation of the pro-mitotic NF-kappaB system.^[16,17] These various molecular changes drive cellular proliferation. Nonetheless, *Cryptosporidium* could have pro or anti apoptotic effects depending the stage of its life cycle.^[16,17,18] At its trophozoite stage it inhibits apoptosis, while it promotes apoptosis at the sporozoite and merozoite stages.^[16]

Toxoplasma gondii

Toxoplasma gondii is a coccidian unicellular protozoan of the class Sporozoa and genus *Toxoplasma*. The first recorded relationship between this obligate intracellular parasite and neoplasia was in 1967.^[5] A positive correlation between the prevalence of *Toxoplasma gondii* and brain cancer in adults has

been demonstrated in 37 countries, while in France, an increased seroprevalence of *Toxoplasma gondii* has been shown to be associated with increased brain cancer mortality.^[9, 19] While elevated levels of anti *Toxoplasma gondii* antibodies have been seen in cases of meningioma and glioma, El-Gayar also documented increased risks for Hodgkin's and intraocular lymphomas in patients infected with the protozoan.^[5, 19,20] In Korea, the incidence of both primary and metastatic brain tumors were observed to be significantly higher in individuals infected with this intracellular protozoan.^[21] In addition, the Chinese study by Yuan et al demonstrated an increase in the incidence of Nasopharyngeal and rectal carcinomas in the presence of Toxoplasmosis.^[22]

Toxoplasma persists in brain tissue within neurons, macrophages and pseudocysts, thus triggering a chronic inflammatory state. It is also known to alter host cell signaling, motility and morphology.^[23] Molecular studies have demonstrated that *Toxoplasma* turns host cells resistant to apoptosis by impairing activation of pro-apoptotic molecules such as caspase-8, Bax and Bak while promoting the actions of anti-apoptotic agents Bcl2 and Bfl1.^[24] It sequesters the inhibitor component of NFkB, thus unleashing its pro-mitotic functions.^[25] This protozoan has also been demonstrated to gain control of host cells, and up regulates miR-1792 cluster which is associated with brain cancers.^[25] This cluster of micro-RNAs is pivotal to the regulation of the cell-cycle, proliferation and apoptosis.^[26] Its dysregulation has been equally strongly linked to haematopoietic and solid cancers, insomuch that it is equally referred to as "oncomir-1".^[26] Cells infected with *Toxoplasma* have been shown to be resistant to Fas-dependent and Fas-independent pathways of Cytotoxic T cell induced apoptosis.^[24] Bradyzoites of this organism have been demonstrated to impede apoptosis following exposure to UV and gamma irradiation, toxins and deficiency of growth factors.^[27-29] The growth effects of *Toxoplasma* have been demonstrated to vary, depending on its developmental stage. Latent slow growing bradyzoites induce gene dysregulation to a lesser extent than faster growing tachyzoites.^[23]

Trichomonas vaginalis

This protozoan is a pear shaped flagellated organism of the class Mastigophora, and genus *Trichomonas* is known to cause vaginitis in females and urethritis in males.^[12] It has recently been linked to two malignancies of the reproductive tract, cervical cancer in females and prostate cancer in males. In the United states and Canada, the incidence of cervical cancer was said to be three times higher in persons with infection

with *T.vaginalis* while infection with the organism was observed in 4-5% of cervical cancer cases in China.^[5, 30] Seropositivity for *T.vaginalis* was associated with a statistically significant elevation in the risk of metastatic cancer of the prostate.^[31,32] African Americans who have the highest risk of carcinoma of the prostate, equally have the highest incidence of trichomoniasis, while the level of serum antibodies against a-actinin protein [derived from *T.Vaginalis*] correlates with an increased risk of having prostate cancer.^[23, 31]

Upon adherence and entry into Prostatic Epithelial cells [PECs], this organism triggers proto-oncogenes such as [c-myc, PIM1 and HMGA1], which drive limitless cell proliferation.^[5] *T.vaginalis* has also been demonstrated to alter the expression of junctional proteins such as E-cadherin, Occludin, and ZO-1.^[5] A *T.vaginalis* protein, homologous to Human Macrophage Inhibitory Factor [MIF] has been identified in infected cells.^[32] This protein, called TvMIF triggers pathways involved in cell proliferation and inflammation. TvMIF activates the anti-apoptotic Akt pathway via silencing of the pro-apoptotic BAD protein.^[32] Its human homologue has been implicated in oncogenic transformation, as over expression has been observed in various human cancers. Interestingly, MIF over expression interferes with p53 activity, thus promoting accumulation of oncogenic mutations.^[33]

With regards cervical cancer, a study by Donders observed that *T. Vaginalis* infection correlated positively with both low risk and high risk HPV infection.^[34] Infection with this protozoan was associated with a slightly higher rate of Atypical Squamous Cells of Undetermined Significance [ASCUS] when co-existing with High Risk HPV than when only the viral infection is present.^[34] As only 1% of HSIL cases seen in this study harbored the protozoan, it's unlikely that the organism has a significant role in its pathogenesis.^[34]

Theileria spp

Theileria is a genus of parasitic protozoan that belongs to the phylum Apicomplexa and is closely related to *Plasmodium*. This intracellular protist is known to cause significant disease and death in both man and cattle, in Africa and Asia.^[23] *Theileria* incites reversible transformation of leucocytes, and induces lymphoproliferative diseases, which is often lethal.^[35]

Theileria is equally known to induce the inhibition of apoptosis. It effects its anti apoptotic effect via the activation of NF-kb sequestering [and thus inhibiting] the TP53 protein.^[23,35, 36] It equally enhances the secretion

of Granulocyte Monocyte-Colony Stimulating Factor [GM-CSF] which stimulates host cell proliferation. This cytokine [GM-CSF] induces the oncogene c-Myc, while constitutive activation of the oncogene C-jun kinase has also been documented.^[37-39] *Theileria* spp has also been known to alter host cell cytoskeleton, increasing the motility of host cells, causing them to behave as leukocyte metastasis.^[23]

Blastocystis hominis

Blastocystis Hominis is a protozoan intestinal parasite belonging to the *Blastocystis* genus of Stramenopiles- vast array of organisms including brown algae, water molds, and diatoms. It has a widespread geographic distribution and is found in countries of all income levels across the world.^[40] This genus of single celled parasites is the most common protozoan infection in the United States. Infection rates vary from about 23% in the States to up to 100% in less developed nations.^[41] It's a very common cause of GI symptoms in cancer patients and the immunosuppressed.^[42] A study carried out in Malaysia observed that 21.08% of patients with colorectal cancer were positive for *Blastocystis* infection.^[43]

Blastocystis sp has been demonstrated to facilitate the growth of colorectal cancer cells in-vitro.^[43] This promitotic impact is mediated through two modalities. Firstly, cells of a colorectal carcinoma express higher levels of Interleukins 6 and 8 in the presence of *Blastocystis*. Signalling induced by these cytokines lead to increased cell turnover and impaired apoptosis.^[42] In the same vein, dysregulation of the p 53 gene and an impaired immune response [due to altered secretion and action of Interferon gamma], contribute to growth of colonic cancer cells.^[43]

Plasmodium falciparum

Alongside Epstein Barr Virus infection, holoendemic malaria, caused by *Plasmodium falciparum* infection, has strong but unproven links to Burkitt lymphoma.^[44] Recently, Cysteine rich interdomain region 1a[CDR-1a], a microbial protein found on the surface of infected erythrocytes has been demonstrated to interact with B lymphocytes. This induces B lymphocyte proliferation, cytomegaly, and secretion of immunoglobulins.^[45,46] This molecule has also been documented to protect B cells from apoptosis.^[45]

NEW ROLES FOR ESTABLISHED PARASITIC CARCINOGENS

Schistosoma haematobium

Its long standing links to the causation of Squamous cell carcinoma of the bladder notwithstanding, *Schistosoma haematobium* has recently been linked to cancers of the prostate and squamous cell carcinoma of

the cervix.^[47-50] Further insight into the oncogenic potential of this trematode has emerged, as a number of studies have uncovered key molecular events such as activation of the H-Ras oncogene and inactivation the tumour suppressors p53 and Retinoblastoma [Rb] genes have been observed in cases of Schistosomiasis associated with cancer of the bladder.^[51,52,53]

Schistosoma mansoni and Schistosoma japonicum

Although *Schistosoma haematobium* appears to be the only member of this specie that has an established role in cancer formation. The documented link between infection by this trematode and squamous cell carcinoma of the bladder has affirmed its status as a carcinogen. Interestingly, new insights have merged on the relationship between other members of this specie and cancers. Fifty one [51%] of cases of Hepatocellular Carcinoma [HCC] seen in Japan, had infection with *Schistosoma japonicum*.^[54] A post mortem study involving 571 autopsies demonstrated an increased incidence of Schistosomiasis among cases of HCC.^[54] This malignancy appears early and in larger numbers in experimental S.jap infection.^[55] In China, a case control analysis showed a strong association between *Schistosoma japonicum* infection and rectal carcinoma but no association with Colonic Carcinoma.^[55]

There have been isolated case reports linking *Schistosoma mansoni* and cancer of the prostate, whereas the organism also has a direct association with HCC. A study of 1577 spleen biopsies from patients with *Schistosoma mansoni* infection showed the presence of follicular lymphomas.^[6] Patients with *Schistosoma japonicum* have higher rates of infection with Hepatitis B and Hepatitis C viruses. This can be partly explained by the increased use of parenteral medication and blood transfusions. In addition cell mediated immunity in active *Schistosoma japonicum*, thus promoting chronic infection by these viruses.^[55] Mostafa in Egypt, observed a direct relationship between chronic Schistosomiasis and Hepatocellular carcinoma, that wasn't restricted to its potentiating of viral hepatitis.^[56]

Altered expression of the tumour suppressor protein p53 in cases of colorectal cancer associated with schistosomal colitis indicates that it is an early/inciting event in the development of colorectal cancer.^[54,57] *Schistosoma mansoni* impairs the functions of cytochrome enzymes, cytoP450, cyto b-5 and NADPH cyto reductase activity, all leading to enhanced effects of aflatoxin on the liver.^[6]

Opisthorchis viverrini

Opisthorchis viverrini, known to be a causative agent of bile duct cancer, has been demonstrated to secrete a

protein, Granulin[Ov-GRN-1], which is reported to be the only helminth induced growth factor known to induce proliferation of mammalian cells.^[58,59] Its human homologue, Human GRN is known to inhibit apoptosis, induce angiogenesis, tumour invasion and anchorage independence.^[59,60]

Taenia solium

Neurocysticercosis is the most common parasitic infection of the central nervous system.^[61] Caused by the tapeworm *Taenia solium*, it has been associated with gliomas of the central nervous system. The association between neurocysticercosis and gliomas has been mostly reported in endemic regions.^[61, 62] A number of studies have demonstrated direct and indirect links between neurocysticercosis and gliomas. It has been hypothesized that the chronic inflammation induced by the parasite may cause cellular proliferation and increases the risk of mutations. Conversely, some authorities have suggested that the neurocysticercosis was a consequence rather than a cause of the formation of gliomas.^[61, 62] This disproved hypothesis was hinged on the suggestion that the immunosuppressed state induced by the cancer would promote the growth of cysticerci.^[63,64]

CONCLUSION

With increased sensitivity and specificity of laboratory techniques and the consequent proficiency in demonstrating alteration of cell proliferation and apoptosis, the role of protozoa in cancer formation has been established. Inasmuch as this could partly explain the sustained increase in the number of malignancies seen in Sub Saharan Africa and other tropical regions of the world, it equally signals the need to screen for cancer associated parasites and treat/give prophylaxis.^[7]

The multi-step nature of microbial/protozoan carcinogenesis provides ample opportunities for interventions to mitigate/prevent cancer.^[65] Antimicrobial treatments have a favourable effect on prognosis in cancers and Oncologists will benefit from an understanding of the mechanisms of infected related carcinogenesis to develop novel effective cancer control strategies. In the same vein, a much more robust investment of resources should be made in the screening and treatment of chronic parasitic infection, as well as a heightened index of suspicion of incipient neoplastic transformation in a background of such infections.

Conflict(s) of interest:

The authors hereby declare that there are no conflicts of interest regarding the publication of this paper

REFERENCES

1. IARC working group on the evaluation of carcinogenic risks to humans. Schistosomes, liver flukes and *Helicobacter pylori*. IARC Monogr Eval Carcinog Risks Hum. 1994;61:1-241.
2. Orem J, Mbidde EK, Lambert B, de Sanjose S, Weiderpass E. Burkitt's lymphoma in Africa, a review of the epidemiology and aetiology. *Afr Health Sci*. 2007;7(3):166-175.
3. CanTreat International. Scaling up cancer diagnostics in developing countries: what we can learn from the HIV/AIDS epidemic. *Ann Oncol*. 2010;21(4):680-682.
4. American Cancer Society. Can Infections lead to cancer. Available from: www.cancer.org. [updated 4/47/2015; cited 14/12/2015]
5. El-Gayar, Mahmoud MM. Protozoa and carcinogenesis. *Parasitol United J*. 2014;7:80-85.
6. Benamrouz S, Conseil V, Creusy C, Calderon E, Decais E, Certad G. Parasites and malignancies, A Review, with emphasis on digestive cancer induced by *Cryptosporidium Parvum* (Alveolata: Apicomplexa). *Parasite*. 2012;19:101-115.
7. Shanju S. Parasites and cancer-A molecular insight. *Glob J Parasitol*. 2015;1:1-7.
8. Kumar V, Abbas A, Aster JC. Neoplasia. In: Robbins and Cotran Pathologic basis of disease [Internet] Canada: Elsevier. 9th. [266-340]
9. Alibek K, Kakpenova A, Baiken Y. Role of infectious agents in the carcinogenesis of brain and head and neck. *Infect Agent Cancer*. 2013;8(7):1-9.
10. Maeda H. Carcinogenesis via microbial infection. *Gan To Kagaku Ryoho* 1998;25(10):1474-1485.
11. Peterson RM, Weidener N. Gastrointestinal neoplasia associated with bowel parasitosis: Real or imaginary? *J Of Trop Med*. 2011;2011.doi:10.1155/2011/234254.
12. Mahan SC. Protozoans. In Gladwin M, Trattler B. *Clinical Microbiology made ridiculously simple*. 4th Edition; 2007; Miami; MedMaster. 324-349
13. Abdou AG, Harba NM, Affi AF, Elnaidany NF. Assessment of *Cryptosporidium parvum* infection in immunocompetent and immunocompromised mice and its role in triggering intestinal dysplasia. *Int JO Inf Dis*. 2013;17:593-600.
14. Sulzyc-Bielicka V, Kuzna-Grygiel W, Kolodziejczyk L, Bielicki D, Kladny J, Stepien-Korzonek M et al. *Cryptosporidiosis* in patients with colorectal cancer. *J Parasitol*. 2007;93(3):722-724.
15. Certad G, Ngouanesavanh T, Guyot K, Gantois N, Chassat T, Mouray A et al. *Cryptosporidium parvum*, a potential cause of colonic adenocarcinoma. *Infect Agent Can*. 2007;2(22).
16. Mele R, Morales MAG, Tosini F, Pozio E. *Cryptosporidium parvum* at different

developmental stages modulates host cell apoptosis in vitro. *Infect Immun.* 2004;72(10):6061-6067.

17. Liu J, Deng M, Lancto CA, Abrahamsen MS, Rutherford MS, Enomoto S. Biphasic modulation of apoptotic pathways in *Cryptosporidium parvum* infected human intestinal epithelial cells. *Infect Immun.* 2009;77(2):837-849.
18. Chen XM, Levine SA, Splinter PL, Tietz PS, Ganong AL, Jobin C et al. *Cryptosporidium parvum* activates nuclear factor kappa B in biliary epithelia, preventing apoptosis. *Gastroenterology.* 2001;120(7):1774-17783.
19. Thomas F, Lafferty KD, Brodeur J, Elgeuro E, Gauthier-Clerc M, Misse D. Incidence of adult brain cancers is higher in countries is higher in countries where the parasite *Toxoplasma gondii* is common. *Biol Lett* 2012; 8:101-103.
20. Ryan P, Hurley SF, Johnson AM, Salzberg M, Lee MW, North JB et al. Tumours of the brain and the presence of antibodies to *Toxoplasma gondii*. *Int J Epidemiol.* 1993;22:412-419.
21. Jung BK, Song H, Kim MJ, Cho J, Shin EH, Chai JY. High *Toxoplasma gondii* seropositivity among brain tumor patients in Korea. *Korean J Parasitol.* 2016;54(2):201-204.
22. Yuan Z, Gao S, Liu Q, Xia X, Liu X, Liu B, et al. *Toxoplasma gondii* antibodies in cancer patients. *Cancer Lett.* 2007;254:71-74.
23. Silmon de Monerri NC, Kim K. Pathogens hijack the epigenome A New Twist on Host-Pathogen Interactions. *Am J Pathol.* 2014;184:897-911.
24. Thirugnanam S, Rout N, Gnanasekar M. Possible role of *Toxoplasma gondii* in brain cancer through modulation of host micro RNAs. *Infect Agent Cancer.* 2013;8:8.
25. Zeiner GM, Norman KL, Thomson JM, Hammond SM, Boothroyd JC. *Toxoplasma gondii* infection specifically increases the levels of key host MicroRNAs. *PloS ONE.* 2010;5(1).
26. Mogilyansky E, Rigoustos I. The mir 17/92 cluster: a comprehensive update on its genomics, genetics, functions and increasingly important and numerous roles in health and disease. *Cell Death Differ.* 2013;20:1603-1614.
27. Laliberte J, Carruthers VB. Host cell manipulation by the human pathogen *Toxoplasma gondii*. *Cell Mol Life Sci.* 2008;65(12):1900-1915.
28. Carmen JC, Hardi L, Sinai AP. *Toxoplasma gondii* inhibits ultraviolet light induced apoptosis through multiple interactions with the mitochondrion-dependent programmed cell death pathway. *Cell Microbiol.* 2006;8(2): 301-315.
29. Nash PB, Purner MB, Leon RP, Clarke P, Duke RC, Curiel TJ. *Toxoplasma gondii* infected cells are resistant to multiple inducers of apoptosis. *J Immunol.* 1998; 160:1824-1830.
30. Sayed el-Ahl SA, el-Wakil HS, Kamel NM, Mahmoud MS. A Preliminary study on the relationship between *Trichomonas vaginalis* and cervical cancer in Egyptian women. *J Egypt Soc Parasitol.* 2002;32(1):167-168.
31. Sutcliffe S, Naece C, Magnuson NS, Reeves R, Alderete JF. Trichomoniosis, a common curable STI and Prostate carcinogenesis-A proposed molecular mechanism. *PLoS Pathog.* 2012;8(8).
32. Twu O, Donders D, Vu A, Mercer F, Stevens GC, de Miguel N, et al. *Trichomonas vaginalis* homolog of macrophage migration inhibitory factor induces prostate cell growth invasiveness and inflammatory responses. *PNAS.* 2013;111(22):8179-8184.
33. Fingerle-Rowson G PO. MIF coordinates the cell cycle with DNA damage checkpoints. Lessons from knockout mice models. *Cell Division.* 2007;2(22).
34. Donders GGG, Depuydt CE, Bogers JP, Vereecken AJ. Association of *Trichomonas vaginalis* and Cytological Abnormalities of the Cervix in Low Risk Women. *PloS ONE.* 2013;8(12):e86266.
35. Dobbelaere D, Heussler V. Transformation of leukocytes by *Theileria parva* and *T. annulata*. *Annu Rev Microbiol.* 1999;53:1-42.
36. Heussler VT, Machado JJ, Fernandez PC, Botteron C, Chen CG, Pearse MJ et al. The intracellular parasite *Theileria parva* protects infected T cells from apoptosis. *Proc Natl Acad Sci U S A.* 1999;96:7312-7.
37. Lizundia R, Chaussepied M, Huerre M, Werling D, Di Santo JP, Langsley G. c-Jun NH2-Terminal Kinase/c-Jun Signaling Promotes Survival and metastases of B lymphocytes transformed by *Theileria*. *Cancer Res* 2006;66(12):6105-6110.
38. Dessauge F, Hilaly S, Baumgartner M, Blumen B, Werling D, Langsley G. c-Myc activation by *Theileria* parasites promotes survival of infected B-lymphocytes. *Oncogene.* 2005;24:1075-83.
39. Dessauge F, Lizundia R, Baumgartner M, Chaussepied M, Langsley G. Taking the Myc is bad for *Theileria*. *Trends Parasitol.* 2005;21(8):377-85.
40. *Blastocystis homini*. available at www.web.stanford.edu/group/parasites2010
41. Amin OM. Seasonal prevalence of intestinal parasites in the United States during 2000. *Am J Trop Med Hyg.* 2002;66(6):799-803.
42. Kumarasamy V, Roslani AC, Rani KU, Govind SK. Advantage of using colonic washouts for *Blastocystis* detection in colorectal cancer patients. *Parasites and Vectors.* 2014;7:1-5.
43. Salomao JF PM, da Silva AR, Leibinger RD, Bellas AR, Campos JM et al. Positive reaction for cysticercosis and multicentric anaplastic oligoastrocytoma *Childs Nerv Syst.* 2006;22:182-5.
44. Moormann AM, Snider CJ, Chelimo K. The

- company malaria keeps: how co-infection with Epstein Barr virus leads to endemic Burkitt lymphoma. *Curr Opin Infect Dis.* 2011;24(5):435-41.
45. Chene A, Donati D, Guerreiro-Cacais AO, Levitsky V, Chen Q et al. A Molecular link between Malaria and Epstein Barr Virus Reactivation. *PLoS Pathog.* 2007;3(6).
 46. Donati D, Zhang LP, Chen Q, Chene A, Flick K, Nystrom M et al. Identification of a polyclonal B-cell activator in *Plasmodium falciparum*. *Infect Immun.* 2004;72(9):5412-8.
 47. Basilio-de-Oliveira CA, Aquino A, Simon EF, Eyer-Silva WA. Concomitant Prostatic Schistosomiasis and Adenocarcinoma: a case report and review. *Braz J Infect Dis.* 2002;6(1):45-9.
 48. Swai B, Poggensee G, Mtweve S, Krantz I. Female genital schistosomiasis as an evidence of a neglected cause of reproductive ill-health: a retrospective histopathological study from Tanzania. *BMC Infect Dis.* 2006;6(134).
 49. Manasseh AN, Echejoh G, Tanko MN, Silas OA, Dakum NK, Mandong BM. Prostatic adenocarcinoma coexisting with Schistosomiasis: A case report and review of literature. *Int J Med Med Sci.* 2009;1(3):33-37.
 50. Helling-Giese G, Sjaastad S, Poggensee G, Kjetland EF, Richter J, Chitsulo L et al. Female genital schistosomiasis[FGS]: relationship between gynaecological and histopathological findings. *Acta Trop.* 1996;62(4):257-67.
 51. Ramchurren N, Cooper K, Summerhayes IC. Molecular events underlying Schistosomiasis-related bladder cancer. *Int J Cancer.* 1995;62:237-44.
 52. Sidransky D, Von Eschenbach A, Tsai YC. Identification of p53 gene mutations in bladder cancers and urine samples. *Science.* 1991;252:700-9.
 53. Ishikawa J, Xu HJ, Hu XS, Yandell DW, Maeda S, Kamidono S et al. Inactivation of the Retinoblastoma gene in human bladder cancer and renal-cell carcinomas. *Cancer Res.* 1991;51:5736-43.
 54. Palumbo E. Association between Schistosomiasis and Cancer. *Infect Dis Clin Pract.* 2007;15:145-8.
 55. Khurana S, Dubey ML, Malla N. Association Of Parasitic Infections and Cancers. *Ind J Micro.* 2005;23(2):74-9.
 56. Mostafa YNA. *Schistosoma mansoni* infection and hepatocellular carcinoma in Egypt. MSc Thesis. Cairo: Cairo University;2012.
 57. Madbouly KM, Senagore AJ, Mukerjee A, Hussien AM, Shehata MA, Navine P et al. Colorectal cancer in a population with endemic *Schistosoma mansoni*: is this an at-risk population? *Int J Colorectal Dis.* 2007;22:175-81.
 58. Smout MJ, Laha T, Mulvenna J, Sripa B, Suttiaprapa S, Jones A, et al. A Granulin-Like Growth Factor Secreted by the Carcinogenic Liver Fluke, *Opisthorchis viverrini*, Promotes Proliferation of Host Cells. *PLoS Pathog.* 2009;5(10):e1000611.
 59. Smout MJ, Mulvenna JP, Jones MK, Loucas A. . Expression, refolding and purification of Ov-GRN-1, a granulin-like growth factor from the carcinogenic liver fluke, that causes proliferation of mammalian cells. *Protein Expr Purif.* 2011;79(2):263-70.
 60. Bateman A, Bennet HP. Granulins: the structure and function of an emerging family of growth factors *J Endocrinol.* 1998;158(145-151).
 61. Kumar N, Bhattacharya T, Kumar R, Radotra BD, Mukherjee KK, Kapoor R, et al. Is Neurocysticercosis a risk factor for glioblastoma multiforme or a mere coincidence. *J Neurosci Rural Pract.* 2013;4:67-9.
 62. Del Brutto OH, Dolezal M, Castillo PR, Garcia HH. Neurocysticercosis and oncogenesis. *Arch Med Res.* 2000;31(2):151-5.
 63. Sanz CR. Host Response in childhood neurocysticercosis. *Childs Nerv Syst.* 1987;3:2067.
 64. Soto-Hernandez JL, Ostrosky-Zeichner L, Tavera G, Gomez-Avina A. Neurocysticercosis and HIV infection: Report of two cases and review. *Surg Neurol.* 1996;45:5761.
 65. Blaser MJ. Understanding Microbe-Induced Cancers. *Cancer Prev Res.* 2008;1(1):15-20.